

Inherited Genetic Markers for Thrombophilia in Northeastern Iran (a Clinical-Based Report)

Fatemeh Keify¹, Mohsen Azimi-Nezhad^{1,2,3}, Narges Zhiyan-abed^{1,4},
Mojila Nasseri¹, Mohammad Reza Abbaszadegan^{*1,5}

Abstract

Background: Thrombophilia is a main predisposition to thrombosis due to a procoagulant state. Several point mutations play key roles in blood-clotting disorders, which are grouped under the term thrombophilia. These thrombophilic mutations are methylenetetrahydrofolate reductase (MTHFR, C677T, and A1298C), factor V Leiden (G1691A), prothrombin gene mutation (factor II, G20210A), and plasminogen activator inhibitor (PAI). In the present study, we assessed the prevalence of the above thrombophilia markers in patients with recurrent pregnancy loss or first and second trimester abortions, infertility, and failed in vitro fertilization (IVF).

Methods: This study was conducted among 457 cases those were referred to detect the inherited genetic markers for thrombophilia. Markers for MTHFR, Factor II, and Factor V were assessed by polymerase chain reaction restriction fragment length polymorphism (PCR-RFLP), and PAI was assessed by Amplification Refractory Mutation System (ARMS-PCR).

Results: Two hundred sixty cases (56.89%) were diagnosed as having at least one thrombophilia marker, whereas 197 cases (43.11%) had no thrombophilia markers and were normal.

Conclusion: According to the current study, the pattern of abnormal genetic markers for thrombophilia in northeastern Iran demonstrates the importance of genetic evaluations in patients who show clinical abnormalities with recurrent spontaneous abortion (RSA) or other serious obstetric complications.

Keywords: Factor II, Factor V, Thrombophilia, MTHFR, PAI, Thrombophilic markers

Introduction

Thrombophilia (hypercoagulability or prothrombotic state) is an abnormality in blood coagulation that increases the risk of thrombosis (1). Such abnormalities can be recognized in 50% of people who have thrombotic episodes (2). Thrombophilia is the tendency to develop thromboses due to inherited defects in the coagulation system. A significant proportion of the population has detectable abnormalities, but most people develop thromboses in the presence of additional risk factors.

Women with thrombophilia are at risk for pregnancy loss and other serious obstetric complications. Recently, studies have focused on thrombophilic factors associated with pregnancy complications. The four most common genetic markers for thrombophilia are; factor V Leiden (FVL, G1691A), methylene tetrahydro folate reductase mutations (MTHFR, C677T and A1298C), prothrombin gene mutation (FII, G20210), and plasminogen activator inhibitor 1 (PAI-1) (3).

1: Pardis Clinical and Genetics Laboratory, Mashhad, Iran.

2: Department of Medical Genetics, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran.

3: Université de Lorraine, Unité de Recherche "Interactions Gène-Environnement en Physiopathologie Cardio Vasculaire" l'UMR INSERM U 1122, IGE-PCV, Nancy, France.

4: Razavi's Social Welfare Organization, Mashhad, Iran.

5: Division of Human Genetics, Immunology Research Center, Avicenna Research Institute, MUMS, Mashhad, Iran.

*Corresponding author: Mohammad Reza Abbaszadegan; Tel: +98 5117112343; Fax: +98 5117112343; E-mail: abbaszadeganmr@mums.ac.ir

Received: Nov 03, 2013; Accepted: Dec 11, 2013

In FVL, at amino acid residue 506, arginine is substituted by glutamine. Due to this substitution factor V becomes resistant to degradation by activated protein C, increasing the risk of venous thromboembolism. In factor II, at position 20210 of the 3' untranslated region (UTR) of the factor II gene, a G to A transition has been found to be associated with increased prothrombin levels and increased risk for venous thrombosis. The homozygous state for the C to T transition at position 677 and A to C transition at position 1298 of MTHFR gene is associated with hyperhomocysteinaemia, which predisposes the carrier to thrombosis (4).

Plasminogen activator inhibitor-1 (PAI-1) is a glycoprotein with a molecular weight of approximately 50 kDa and a member of the serine protease inhibitor super-family. PAI-1 is a rapid and specific inhibitor of both tissue-type plasminogen activator and urokinase, and is considered as the primary regulator of plasminogen activation *in vivo*. Abnormally high concentrations of PAI-1 in blood observed in atherothrombotic and other diseases seem to result from changes in the rate of PAI-1 gene expression in tissues rather than from release of stored PAI-1 in the cells. The 4G/5G polymorphism in the PAI-1 gene promoter seems to be one of DNA sequence variation, which has functional importance in regulating expression of the PAI-1 gene. This polymorphism, located 675 base pairs upstream of the PAI-1 gene transcription start site, is characterized by a single guanosine deletion/insertion, resulting in two alleles containing either 4 or 5 consecutive guanosines.

Our ability to detect genetic abnormalities responsible for thrombotic propensity has improved in recent years. However the evidence for the role of inherited thrombophilias such as factor V Leiden, prothrombin G20210A mutation, the methylenetetrahydrofolate reductase (C677T, A1298T) mutation, and the plasminogen activator inhibitor is less clear.

As it is conceivable that thrombophilias were diagnosed in these series due to thromboembolic phenomena, we considered it necessary to assess the prevalence of thrombophilias in patients with recurrent miscarriages and no previous thrombophilic phenomena. In the present study, we assessed the prevalence of the above thrombophilia markers in patients with recurrent pregnancy losses or first and

second trimester abortions, infertility, and failed *in vitro* fertilization (IVF).

Materials and Methods

In this study, over a three years period (2011-2014), 468 cases were attended by specialists in the Genetic Counseling Services in the Pardis Clinical and Genetics Laboratory (PCGL) or other facilities.

Genomic DNA was extracted from peripheral blood samples by the salting out method (5). Samples with optical density (OD) 260nm/280 ratios of 1.5 – 1.8 and final concentrations of 5 ng/μl were used for amplifications.

The G1691A polymorphism in factor V Leiden was detected by PCR amplification of a 206 bp fragment and *MnII* digestion, as previously described (6). The C677T and A1298T substitutions in the MTHFR gene were identified using *HinfI* and *MboII* cleavage of 494 and 237bp PCR products, respectively (7). For identification of the G20210A substitution in the factor II gene, a 345bp fragment from the 3' UTR was amplified by PCR using the same primers as described (8) and digested with *HindIII*. For PAI, Amplification Refractory Mutation System (ARMS-PCR) was used to detect the 4G/5G polymorphism.

Statistical analysis was performed using SPSS version 16. The genotype distributions of each mutation and their frequencies were compared between patients.

Results

Two hundred sixty nine patients (57.48%) were diagnosed with at least one thrombophilia marker, whereas 199 patients (42.52%) had no thrombophilia markers and were normal. The prevalence of the thrombophilias assessed is shown in Table 1.

Table 1. Frequency of thrombophilia markers in subjects.

Thrombophilia Marker	Frequency	
	Normal	Abnormal
MTHFR	29	244
Factor II	157	4
Factor V	12	4
PAI	1	17
Total	199 (42.52%)	269 (57.48%)

Figure 1 shows some of amplified and digested products for MTHFR, Factor II, Factor V, and PAI

following agarose gel electrophoresis.

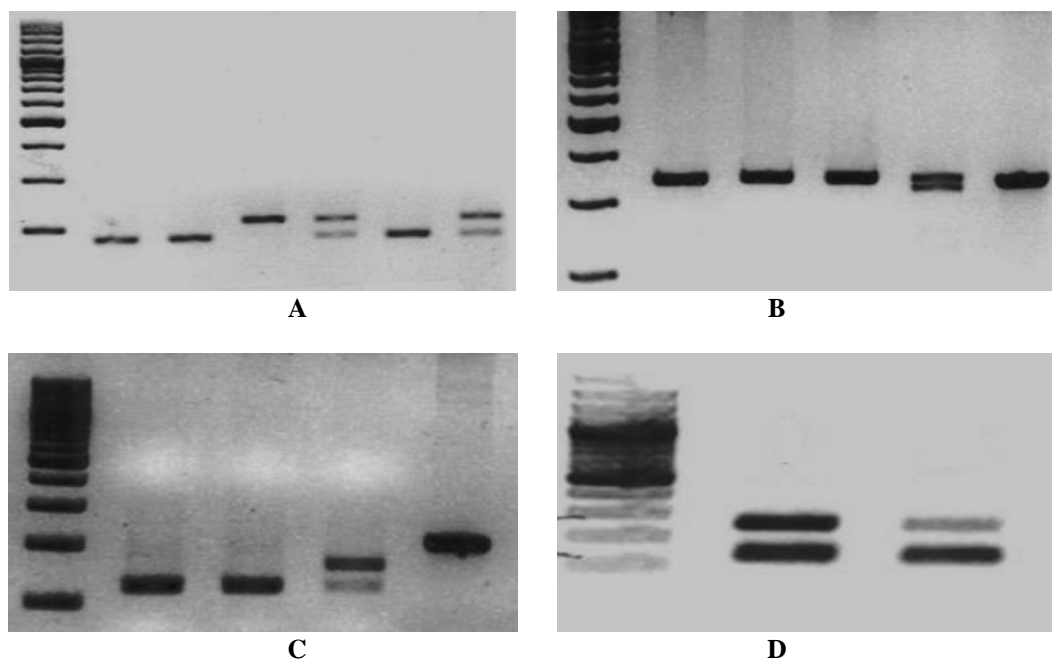


Fig. 1. Agarose gel electrophoresis of samples from thrombophilia patients; (A) Analysis of the A1298C polymorphism in the MTHFR gene. The samples were PCR-amplified and digested with MboII. Samples in lanes 1, 2, and 5 came were isolated from normal subjects. Samples in lanes 4 and 6 were from subjects who were heterozygous, and the sample in lane 3 came from a subject who was homozygous for the A1298C polymorphism. (B) Analysis of the G20210A polymorphism in Factor II. The samples were PCR-amplified and digested with HindIII.: Lanes 4 is heterozygous; 1, 2 and 3 are normal; and 5 is uncut. Lanes 1, 2, and 3 were isolated from normal subjects. Lane 4 was isolated from a subject who was heterozygous and lane 5 is undigested. (C) Analysis of the G1691A polymorphism in Factor V Leiden. The samples were PCR-amplified and digested with MnlI: The samples in lanes 1 and 2 were isolated from normal subjects. The sample in lane 3 was from a heterozygous subject, and lane 4 is undigested. (D) Analysis of PAI polymorphisms using ARMS-PCR: The samples in lanes 1 and 2 were isolated from subjects with the 4G and 5G polymorphisms, respectively.

MTHFR

The genotype distribution of the MTHFR mutations in subjects and their case presentations are shown in Table 2. Of 273 subjects analyzed, 29 (10.62%) were normal for MTHFR.

The heterozygous and homozygous C677T polymorphisms were found in 18.32% (50/273) and 9.52% (26/273) of subjects, respectively. The heterozygous and homozygous A1298C polymorphisms were found in 20.88% (57/273) and 12.45 % (34/273), respectively. The heterozygous C677T and A1298C mutations were found in 28.21% of subjects (77/273).

Factor II

The genotype distribution of the G20210A polymorphism in subjects and the genotype distribution based on RSA, infertility, and failed IVF are shown in Table 3. The frequency of the heterozygous G20210A mutation was 1.91% (4/209) and 98.09% (205/209) had normal genotypes.

Factor V

The genotype distribution of the G1691A mutation in subjects and the genotype distribution based on RSA and infertility shown in Table 4. The heterozygous and homozygous G1691A mutation was found in 5.0 % (2/40) and normal individuals were 90.0% (36/40).

Table 2. Distribution and case presentations for subjects with MTHFR mutations

<i>Mutation</i>	<i>Presenting</i>	<i>Frequency</i>	<i>Percentage %</i>
Heterozygous C677T	RSA	47	18.32
	Infertility	2	
	Failed IVF	1	
	Total	50	
Homozygous C677T	RSA	25	9.52
	Infertility	1	
	Failed IVF	0	
	Total	26	
Heterozygous A1298C	RSA	51	20.88
	Infertility	5	
	Failed IVF	1	
	Total	57	
Homozygous A1298C	RSA	30	12.45
	Infertility	3	
	Failed IVF	1	
	Total	34	
Heterozygous C677T and A1298C	RSA	72	28.21
	Infertility	4	
	Failed IVF	1	
	Total	77	
Normal	RSA	23	10.62
	Infertility	4	
	Failed IVF	2	
	Total	29	
Total		273	100

Table 3. Distribution and case presentations for subjects with factor II (G20210A) mutations.

<i>Mutation</i>	<i>Presenting</i>	<i>Frequency</i>	<i>Percentage %</i>
Heterozygous G20210A	RSA	4	1.91
	Total	4	
Normal	RSA	134	75.60
	Infertility	19	
	Failed IVF	5	
	Total	158	
Normal for factor II, but with MTHFR polymorphisms	Heterozygous C677T	8	22.49
	Homozygous C677T	5	
	Heterozygous A1298C	17	
	Homozygous A1298C	4	
	Heterozygous C677T and A1298C	2	
	Total	11	
Total		209	100

PAI

The genotype distribution of the PAI polymorphism in subjects and the genotype distribution based on RSA and infertility shown in Table5. The frequency

of the heterozygous 4G/5G mutation was 96.77% (30/31) and the frequency of homozygous 5G/5G mutation was 3.23% (1/31).

Table 4. Distribution and case presentations for subjects with Factor V (G1691A) mutations

<i>Mutation</i>	<i>Presenting</i>	<i>Frequency</i>	<i>Percentage %</i>
Homozygous	RSA	1	5.0
	Infertility	1	
	Total	2	
Heterozygous	RSA	1	5.0
	Infertility	1	
	Total	2	
Normal	RSA	12	30.0
	Total	12	
Normal for Factor II and Factor V	RSA	6	42.5
	Infertility	1	
	Total	7	
Normal for Factor V, but with MTHFR polymorphisms	Heterozygous C677T	3	12.9
	Homozygous C677T	3	
	Heterozygous A1298C	5	
	Homozygous A1298C	1	
	Heterozygous C677T and A1298C	5	
	Total	17	
Total		40	100

Table 5. Distribution and case presentations for subjects with PAI mutations.

<i>Mutation</i>	<i>Presenting</i>	<i>Frequency</i>	<i>Percentage %</i>
Heterozygous 4G/5G	RSA	15	51.62
	Infertility	1	
	Total	16	
Homozygous 5G/5G	RSA	1	3.23
Homozygous 4G/4G	-	0	0
Normal	-	0	
Hetero 4G/5G with Factor II	RSA	4	19.35
	Mutation analysis	1	
	Embolism	1	
	Total	6	
Hetero 4G/5G with MTHFR	Heterozygous C677T and A1298C	1	12.9
	Heterozygous A1298C	1	
	Normal	2	
	Total	4	
Hetero 4G/5G with Factor V	RSA	4	12.9
	Total	4	
Total		31	100

Discussion

Several studies have reported that factor V Leiden, which is responsible for more than 75% of inherited activated protein C resistance, is a common thrombotic risk factor affiliated with RPL (10-12). Other studies have reported an association between factor V and late pregnancy loss (13). Goodman CS et al. reported an association between 4G/5G genotype of PAI-1 and recurrent pregnancy loss (14). The relationship of methylenetetrahydrofolate reductase (MTHFR) gene C677T, factor V (FV) gene G1691A and prothrombin (PT) gene G20210A polymorphisms to unexplained recurrent early spontaneous abortion were examined

by Xu et al. They showed that the genetic polymorphisms of MTHFR C677T are associated with spontaneous abortion. These results supported those of previous studies. (15-18). It was reported that there is no increased prevalence of FVL, FII, and MTHFR in recurrent early pregnancy loss (19). The presumed relationship between thrombophilia and recurrent pregnancy loss has become sufficient to allow the presence of thrombophilias to be an indication for treatment with anticoagulant drugs (20).

Acknowledgements

This project was financially supported by Pardis clinical and genetic laboratory. We would like to thank

referral cases of PCGL for contributing in this project.

References

1. Heit JA. Thrombophilia: common questions on laboratory assessment and management. *Hematology Am Soc Hematol Educ Program*. 2007; 1: 127–35.
2. Kyrle PA, Rosendaal FR, Eichinger S. Risk assessment for recurrent venous thrombosis. *Lancet*. 2010 Dec;376(9757):2032–39.
3. D'Uva M, Micco PD, Strina I, Placido GD. Recurrent pregnancy loss and thrombophilia. *J Clin Med Res*. 2010;2(1):18–22.
4. Carp H, Salomon O, Seidman D, Dardik R, Rosenberg N, Inbal A. Prevalence of genetic markers for thrombophilia in recurrent pregnancy loss. *Hum Reprod*. 2002 Jun;17(6):1633–37.
5. Miller SA, Dykes DD, Polesky HF. A simple salting out procedure of extracting DNA from human nucleated cells. *Nucl. Acid Res*. 1988 Feb;16(3):1215–17.
6. Salomon O, Steinberg DM, Zivelin A, Gitel S, Dardik R, Rosenberg N, et al. Single and combined prothrombotic factors in patients with idiopathic venous thromboembolism. *Arterioscler Thromb Vasc Biol*. 1999 Mar;19(3):511–18.
7. Frosst P, Blom HJ, Milos R, Goyette P, Sheppard CA, Matthews RG, et al. A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase. *Nat Genet*. 1995 May;10(1):111–13.
8. Poort SR, Rosendaal FR, Reissma PH, Bertina RM. A common genetic variation in the 3'-untranslated region of the prothrombin gene is associated with elevated plasma prothrombin levels, and an increase in venous thrombosis. *Blood*. 1996 Nov;88(10):3698–703.
9. Pickering W, Marriott K, Regan L. G20210A prothrombin gene mutation: prevalence in a recurrent miscarriage population. *Clin Appl Thromb Hemost*. 2001 Jan;7(1):25–8.
10. Rai R, Shlebak A, Cohen H, Backos M, Holmes Z, Marriott K, et al. Factor V Leiden and acquired activated protein C resistance among 1000 women with recurrent miscarriage. *Hum Reprod*. 2001 May;16(5):961–65.
11. Grandone E, Margaglione M, Colaizzo D, d'Addeda M, Cappucci G, Vecchione G, et al. Factor V Leiden is associated with repeated and recurrent unexplained fetal losses. *Thromb Haemost*. 1997 May;77(5):822–24.
12. Hatzis T, Cardamakis E, Drivalas E, Makatsoris K, Bevan D, Pantos C, et al. Increased resistance to activated protein C and factor V Leiden in recurrent abortions. Review of other hypercoagulability factors. *Eur J Contracept Reprod Health Care*. 1999 Sep;4(3):135–44.
13. Martinelli I, Taioli E, Cetin I, Marinoni A, Gerosa S, Villa MV, et al. Mutation in coagulation factors in women with unexplained late fetal loss. *N Engl J Med*. 2000 Oct;343(14):1015–18.
14. Goodman CS, Coulam CB, Jeyendran RS, Acosta VA, Roussev R. Which thrombophilic gene mutations are risk factors for recurrent pregnancy loss? *Am J Reprod Immunol*. 2006 Oct;56(4):230–36.
15. Xu L, Liu XM, Zhang HY, Zhao J, Qi QW, Chang YF. Relationship between three thrombophilic gene mutations and unexplained recurrent early spontaneous abortion. *Zhonghua Fu Chan Ke Za Zhi*. 2007 Mar;42(3):180–3.
16. Nelen WL, Steegers EA, Eskes TK, Blom JH. Genetic risk factor for unexplained recurrent early pregnancy loss. *Lancet*. 1997 Sep;350(9081):861.
17. Unfried G, Griesmacher A, Weismüller W, Nagele F, Huber CJ, Tempfer CB. The C677T polymorphism of the methylenetetrahydrofolate reductase gene and idiopathic recurrent miscarriage. *Obstet Gynecol*. 2002 Apr;99(4):614–19.
18. Mtraoui N, Zammiti W, Ghazouani L, Braham NJ, Saidi S, Finan RR, et al. Methylenetetrahydrofolate reductase C677T and A1298C polymorphism and changes in homocysteine concentrations in women with idiopathic recurrent pregnancy losses. *Reprod*. 2006 Feb;131(2):395–401.
19. Kutteh WH, Park VM, Deitcher SR. Hypercoagulable state mutation analysis in white patients with early first-trimester recurrent pregnancy loss. *Fertil Steril*. 1999 Jun;71(6):1048–54.
20. Younis JS, Ohel G, Brenner B, Ben-Ami M. Familial thrombophilia- the scientific rationale for thrombophylaxis in recurrent pregnancy loss. *Human Reprod*. 1997 Mar;12(7):1389–90.