

Effect of Treatment with Thyme Extract on Urinary Levels of Melatonin in an Experimental Autoimmune Encephalomyelitis Mouse Model

Merat Mahmoodi¹, Farnaz Sedghy*¹, Abdollah Jafarzadeh¹

Abstract

Background: Thymus vulgaris, or thyme belongs to the *Lamiaceae* family of aromatic plant species and has established antioxidant and anti-inflammatory properties. We examined the association between thyme extract treatment to recovered urinary levels of melatonin, a hormone with neuroprotective effects, in mice induced with EAE.

Methods: Eight B6 mice induced with EAE were randomized into two groups and exposed to either 50 mg/kg of thyme extract or PBS. After EAE induction, mice were injected i.p every other day from day 0 to 21. Four B6 mice without EAE were considered the healthy control group. Urine samples were collected consecutively for two 24 h periods on day 19 and 20. We examined whether thyme extract treatment modified urinary melatonin sulfate concentration (ng/mL) in EAE-induced mice using an ELISA.

Results: The clinical score and body weight in thyme-treated EAE group were significantly lower in comparison to the EAE control group at indicated time points. The urinary melatonin concentration was significantly lower in the EAE control group compared to the healthy mice. There was no significant difference between thyme-treated and EAE groups regarding the urine melatonin concentration.

Conclusions: Our results show that exposing EAE mice to thyme extract improved their clinical symptoms, however, there was no significant effect on urinary melatonin concentration.

Keywords: Enzyme-linked immunosorbent assay (ELISA), Experimental autoimmune encephalomyelitis (EAE), Melatonin, Thymus vulgaris (Thyme), Urine.

Introduction

Multiple sclerosis (MS) is one of the most common inflammatory diseases of the central nervous system (CNS) in young adults. Previous research documents that several factors are involved in MS pathogenesis including neuronal loss due to oxidative stress, and infiltration of immune cells and neuro-immunomodulatory agents such glucocorticoids, opioids, prolactin, and N-acetyl-5methoxy-tryptamine (melatonin) into the CNS (1-4). The melatonin hormone is secreted from the pineal gland and regulates the body's circadian rhythm. Recent studies suggest a neuroprotective and immunomodulatory role for melatonin in inflammatory and autoimmune diseases (2, 4-6). For example, melatonin was found to modulate the

immune response by decreasing T helper type 1 increasing the anti-(Th1) responses and inflammatory/Th1 ratio (7). In addition, Farez et al., found that melatonin suppressed the generation of TH-17 cells and enhanced the levels of interleukin-10 (IL-10) producing type 1 regulatory T (Tr1) cells (8). Moreover, several investigations demonstrate that melatonin levels are inversely correlated with the severity of MS symptoms, such as fatigue, insomnia and depression (1, 5, 9, 10). Interestingly, there is evidence to support that altered levels of melatonin in MS patients can be corrected after with interferon beta treatment (IFN-B) Natalizumab (11). Common treatments improving clinical MS include IFN-β, glatiramer acetate and corticosteroids. However, due to the side effects and inefficacy of these drugs, there is a growing interest in treating MS using herbal therapeutics (12). Thyme, an aromatic plant species belonging to the Lamiaceae family, is known to possess antioxidant and antiinflammatory characteristics (13-17). To the authors' knowledge, there are limited publications discussing the effects of thyme extract in autoimmune diseases. Our previous research demonstrates that thyme extract has anti-inflammatory properties and can improve the clinical outcome of mice induced with experimental autoimmune encephalomyelitis (EAE), a widely used animal model of MS (18). In this study, we examined whether our previous observations correlated to urinary melatonin concentration in EAE induced mice.

Materials and methods

Mice

C57BL/6 (B6) mice (8–12 weeks old females) were purchased from Royan Institute of Isfahan, Iran. Animals were housed under controlled conditions with a 12 h day and 12 h night cycle. All experiments were approved by Animal Ethics Committee of Mashhad University of Medical Sciences, Mashhad, Iran.

EAE induction

Mice were administered 200 µl of emulsion containing 400 µg myelin oligodendrocyte glycoprotein (MOG) 35–55 peptide (SBS Genetech Co. Ltd., Beijing, China) and 100 µl complete Freund's adjuvant (CFA) (Sigma-Aldrich, St. Louis, MO, USA) (0.4 mg of Mycobacterium inactivated tuberculosis) subcutaneously (s.c) between the shoulders and over the flank. All mice received 250 ng pertussis toxin (Sigma-Aldrich, St. Louis, MO, USA) intraperitoneally (i.p) on the day of EAE induction and 48 h later. Clinical signs of animals were evaluated daily and scored using a 0-8 point scale: 0=no symptoms; 1=partial loss of tail tonicity; 2=complete loss of tail tonicity; 3=flaccid tail and abnormal gait; 4=hind leg paralysis; 5=hind leg paralysis with hind body paresis; 6=hind and foreleg paralysis; and 7=death (19).

Study design

Eight B6 mice were induced with EAE and randomized into two groups, four each. EAE mice in the experimental group were exposed to thyme extract (50 mg/kg), and 2% phosphate-buffer saline (PBS) in the control group. Four mice without EAE induction was considered the healthy control group. Both EAE groups were injected i.p from day 0 to 21 every other day and sacrificed on day 22 after MOG immunization, according to the animal ethics protocol.

Extract preparation of Thyme

Thymus vulgaris (Thyme) was collected from Arac Farms (Arac, Iran) during the summer of 2016 and its identity was confirmed by the Herbarium Institute of Kerman, Iran. A hydro-alcoholic extract was prepared by drying the aerial parts of the thyme plant. Next, 100 g of thyme powder was soaked in 70% ethanol and left in the dark shaking (mildly) for 72 h. Then, the liquid phase was filtrated and concentrated in a rotary evaporator at 40°C until a semi-dried extract with 20% of the initial weight of the powder remained. Finally, the extracts were stored at -20°C and resolved in 2% PBS (18).

Urine collection

Mice were placed in cages that were equipped with a beaker under a stainless-steel grid one at a time. Urine was collected in the beaker consecutively over two 24 h periods on days 19 and 20. Samples were stored at −30°C for analysis via ELISA method. The melatonin concentration was calculated as the average of the two days.

Urinary melatonin detection

Melatonin sulfate (6-sulfatoxymelatonin) levels in urine was detected using an ELISA kit (IBL, Hamburg, Germany; cat.no. RE54031) according to the manufacturer's instructions.

Statistical analysis

Data and statistical analyses were performed using GraphPad Prism version 6.01. The data was represented as the mean \pm S.E.M. We used a t-test for parametric data and a Wilcoxon signed-rank test for non-parametric data. Statistical difference is indicated as *, P < 0.05.

Results

Thyme extract treatment improved the clinical score in EAE-induced mice

After MOG immunization, the first clinical signs of EAE appeared on days 9 and 10 in the untreated and thyme-treated group, respectively. The clinical score in thyme-treated mice were significantly lower in comparison to

the EAE control group at indicated time points (P<0.05 at day 10, and P<0.001 at days 13 to 21) (Fig. 1). Moreover, the body weight of mice in the thyme-treated group was significantly higher compared to untreated-EAE mice at the indicated time points (P<0.05 at day 13 and P<0.001 at days 14 to 21) (Fig. 2).

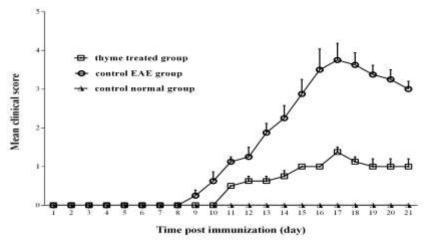


Fig. 1. The mean clinical score in EAE induced mice treated with thyme extract in comparison to the EAE control group (not treated)

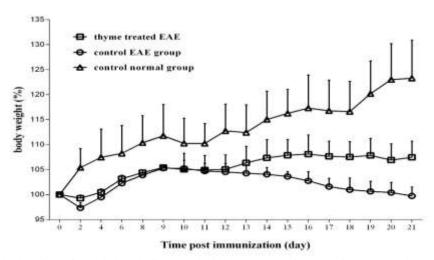


Fig. 2. The body weight of EAE induced mice treated with thyme extract in comparison to control groups (not treated)

No significant difference in urinary melatonin between thyme-treated and untreated-EAE mice

Following thyme extract treatment and clinical assessment, melatonin sulfate concentration in collected urine samples was measured in the thymetreated EAE group, untreated-EAE control and healthy control groups. The average melatonin concentration was significantly higher in normal healthy mice compared to the untreated-EAE control

group (P=0.028). Although melatonin levels were higher in the thyme-treated EAE group compared to the untreated-EAE group, the difference was not found to be statistically significant (P=0.11) (Fig. 3). Lastly, there was no significant difference between the healthy group and the thyme-treated EAE group regarding urinary melatonin concentration.

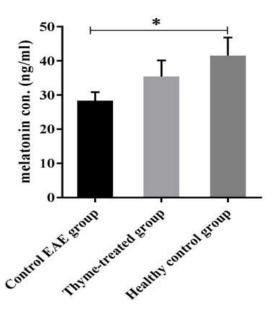


Fig. 3. The average concentration of urine melatonin sulfate levels in EAE induced mice treated with thyme extract in comparison with EAE and healthy control (not treated) groups.

Discussion

Recent studies have indicated various immunomodulatory roles of melatonin in a number of immune-related diseases including autoimmune disorders (4-6). The levels of this hormone are altered during the course of disease and is inversely related to disease severity. Therefore, finding new treatments with the least adverse effects able to modify melatonin levels could be of interest. Melatonin is present in many plants with variable levels (20,21). Therefore, treating autoimmune diseases with these plants may have promising results. This paper aimed to evaluate the effect of thyme extract treatment on melatonin sulfate levels in urine samples of EAE-induced mice. The results of this research showed that despite a decrease in melatonin in EAE-induced mice, there was no significant difference in urinary melatonin sulfate between the thyme-treated and untreated mice.

Evidence in the literature suggests an inverse correlation between the levels of melatonin and the severity of autoimmune diseases. In this study, we observed significantly lower levels of melatonin in EAE-induced control mice compared to the healthy mice. Consistent with these results, Gholipour et al. showed that lower levels of urine 6-sulphatoxymelatonin (aMT6s)

was negatively associated with Functional Composite score (FCS). The FCS factor is used to assess the overall disability impact in MS patients, suggesting a novel method for assessing MS severity (4). Another study demonstrates that there is an inverse relationship between nocturnal melatonin serum levels and major depression, indicating that melatonin deficiency could lead to the development of depression in MS patients (11). In addition, some published papers have shown that there can be negative consequences from disrupting melatonin concentration and the occurrence of MS. For instance, Hedstrom et al. observed that working shift, which causes circadian disruption, could enhance the risk of MS early on (3). Melatonin depletion has also been reported in other autoimmune diseases, for example, a study by Wang et al. showed that decreased plasma melatonin levels in systemic lupus erythematosus (SLE) patients correlated with clinical manifestations (22).

Whether melatonin plays a role in improving clinical symptoms of MS is still a matter of debate. Previous investigations have reported the effect of various drugs and treatments on melatonin levels. In a study conducted on 13 MS and 12 healthy subjects, treatment with IFN-β could ameliorate decreased urinary levels of 6sulphatoxy-melatonin (6-SMT), fatigue and sleep efficiency in MS patients (13). In addition, Natalizumab, one of the most efficient treatments for MS, increased serum melatonin levels as well as antioxidants in 18 patients with relapsing remitting multiple sclerosis (RRMS) (1). Treatment with Luzindole, an antagonist for the melatonin membrane receptor inhibited EAE development in mice following immunization with spinal cord homogenate (23). In contrast, our results found no significant difference in the levels of melatonin sulfate in the urine of thymetreated and untreated mice. This negative result may be attributed to the selected dose of the extract. In this study, we only tested one dose of thyme extract (50 mg/kg), therefore, using a higher dose could potentially alter melatonin levels. Moreover, our findings were limited by a small sample size in each group. Finally, Thymus vulgaris could modulate the immune system

through other pathways not related to melatonin regulation.

In conclusion, these results demonstrated that urinary levels of melatonin were decreased in an EAE disease model. Furthermore, although treatment of EAE-induced mice with thyme extract improved disease progression of

References

- 1. Bahamonde C, Conde C, Aguera E, Lillo R, Luque L, Gascon F, et al. Elevated melatonin levels in natalizumab-treated female patients with relapsing-remitting multiple sclerosis: relationship to oxidative stress. European journal of pharmacology. 2014;730(9):26-30.
- 2. Farhadi N, Oryan S, Nabiuni M. Serum levels of melatonin and cytokines in multiple sclerosis. Biomedical journal. 2014;37(2):90-2.
- 3. Hedstrom AK, Akerstedt T, Hillert J, Olsson T, Alfredsson L. Shift work at young age is associated with increased risk for multiple sclerosis. Annals of neurology. 2011;70(5):733-41.
- 4. Gholipour T, Ghazizadeh T, Babapour S, Mansouri B, Ghaffarpour M, Siroos B, et al. Decreased urinary level of melatonin as a marker of disease severity in patients with multiple sclerosis. Iranian Journal of Allergy, Asthma and Immunology. 2015;14(1):91-7.
- 5. Ghareghani M, Dokoohaki S, Ghanbari A, Farhadi N, Zibara K, Khodadoust S, et al. Melatonin exacerbates acute experimental autoimmune encephalomyelitis by enhancing the serum levels of lactate: A potential biomarker of multiple sclerosis progression. Clinical and Experimental Pharmacology and Physiology. 2017;44(1):52-61.
- 6. Wen J, Ariyannur PS, Ribeiro R, Tanaka M, Moffett JR, Kirmani BF, et al. Efficacy of N-Acetylserotonin and Melatonin in the EAE Model of Multiple Sclerosis. Journal of Neuroimmune Pharmacology. 2016;11(4):763-73.
- 7. Álvarez-Sánchez N, Cruz-Chamorro I, Díaz-Sánchez M, Sarmiento-Soto H, Medrano Campillo P, Martínez-López A, et al. Melatonin reduces inflammatory response in peripheral T helper lymphocytes from relapsing-remitting multiple sclerosis patients. Journal of pineal research. 2017;63(4): e12442.

EAE, it had no significant effect on the urinary melatonin levels.

Acknowledgment

This work was supported by Rafsanjan University of Medical Sciences, Rafsanjan, Iran (grant number 97238).

- 8. Farez MF, Calandri IL, Correale J, Quintana FJ. Anti-inflammatory effects of melatonin in multiple sclerosis. BioEssays. 2016;38(10):1016-26.
- 9. Damasceno A, Moraes AS, Farias A, Damasceno BP, dos Santos LMB, Cendes F. Disruption of melatonin circadian rhythm production is related to multiple sclerosis severity: a preliminary study. Journal of the neurological sciences. 2015;353(1-2):166-8.
- 10. Chen SJ, Huang SH, Chen JW, Wang KC, Yang YR, Liu PF, et al. Melatonin enhances interleukin-10 expression and suppresses chemotaxis to inhibit inflammation in situ and reduce the severity of experimental autoimmune encephalomyelitis. International immunopharmacology. 2016;31(2):169-177.
- 11. Akpinar Z, Tokgoz S, Gokbel H, Okudan N, Uguz, Yulmaz Glm. The association of nocturnal serum melatonin levels with major depression in patients with acute multiple sclerosis. Psychiatry research. 2008;161(2):253-7.
- 12. Jantan I, Ahmad W, Bukhari SNA. Plant-derived immunomodulators: an insight on their preclinical evaluation and clinical trials. Frontiers in plant science. 2015;6:655.
- 13. Melamud L, Golan D, Luboshitzky R, Lavi I, Miller A. Melatonin dysregulation, sleep disturbances and fatigue in multiple sclerosis. Journal of the neurological sciences. 2012;314(1):37-40.
- 14. Yu YM, Chao TY, Chang WC, Chang MJ, Lee MF. Effect of thymol on oxidative stress and inflammation-related gene expression in high fat diet-induced hyperlipidemic rabbits. The FASEB Journal. 2016;30(Suppl. 1):1174.4-1174.4.
- 15.Peterson LK, Fujinami RS. Inflammation, demyelination, neurodegeneration and neuroprotection in the pathogenesis of multiple

sclerosis. Journal of neuroimmunology. 2007;184(1-2):37-44.

16. Babaei M, Abarghoei ME, Ansari R, Vafaei AA, Taherian AA, MM Akhavan, et al. Antispasmodic effect of hydroalcoholic extract of Thymus vulgaris on the guinea-pig ileum. Natural product research. 2008;22(13):1143-50.

17. Meeran MFN, Jagadeesh GS, Selvaraj P. Thymol attenuates inflammation in isoproterenol induced myocardial infarcted rats by inhibiting the release of lysosomal enzymes and downregulating the expressions of proinflammatory cytokines. European journal of pharmacology. 2015;754(9):153-61.

18. Mahmoodi M, Ayoobi F, Aghaei A, Rahmani M, Taghipour Z, Hosseini A, et al. Beneficial effects of Thymus vulgaris extract in experimental autoimmune encephalomyelitis: Clinical, histological and cytokine alterations. Biomedicine & Pharmacotherapy. 2019;109(1):2100-8.

19. Haghmorad D, Mahmoudi MB, Salehipour Z, Jalayer Z, Rastin M, Kokhaei P, et al. Hesperidin

ameliorates immunological outcome and reduces neuroinflammation in the mouse model of multiple sclerosis. Journal of neuroimmunology. 2017;302(1):23-33.

20. Aguilera YM, Rebollo-Hernanz T, Herrera LT, Cayuelas P, Rodríguez-Rodríguez Á, de Pablo ÁLL et al. Intake of bean sprouts influences melatonin and antioxidant capacity biomarker levels in rats. Food & function. 2016;7(3):1438-45.

21. Jana S, Rastogi H. Effects of Caffeic Acid and Quercetin on in Vitro Permeability, Metabolism and in Vivo Pharmacokinetics of Melatonin in Rats: Potential for Herb-Drug Interaction. European journal of drug metabolism and pharmacokinetics. 2017;42(5):781-91. 22. Wang P, Li HM, Zou YF, Tao JH, Pan HF. Plasma melatonin levels do not differ in SLE patients. Zeitschrift für Rheumatologie. 2018;77(1):66-70.

23. Constantinescu C, Hilliard B, Ventura E, Rostami A. Luzindole, a melatonin receptor antagonist, suppresses experimental autoimmune encephalomyelitis. Pathobiology. 1997;65(4):190-4.